

1 **Type of article:** Regular Article

2

3 **Title:** Effects of *Lactiplantibacillus pentosus* ONRICb0240-containing food on Reducing  
4 Immunosuppression in Judo Athletes During Rapid Weight Loss

5

6 **Brief running head:** Lactiplantibacillus and immune function in judo

7

8 **Authors:** Kazuhiro Shimizu<sup>1,\*</sup>, Satoshi Hattori<sup>2,#</sup>, Megumi Matsumoto<sup>3</sup>, Hiroaki  
9 Hiraoka<sup>4</sup>, Yoko Tanabe<sup>1,5,6</sup>, Yukichi Hanaoka<sup>1</sup>, Takashi Ono<sup>4,7</sup>, Fuminori Kimura<sup>4</sup>,  
10 Koichiro Hamada<sup>2</sup>, Koichi Watanabe<sup>4</sup>, Takeshi Kukidome<sup>1</sup>

11

12 **Affiliations:**

13 <sup>1</sup>Department of Sport Science and Research, Japan Institute of Sports Sciences, 3-15-1  
14 Nishigaoka, Kita-ku, Tokyo, 115-0056, Japan

15 <sup>2</sup>Saga Nutraceuticals Research Institute, Otsuka Pharmaceutical Co., Ltd., 5006-5 Aza  
16 Higashiyama, Omagari, Yoshinogari, Kanzaki-cho, Saga, 842-0195, Japan

17 <sup>3</sup>Department of Physical Education, College of Humanities and Sciences, Nihon  
18 University, 3-25-40 Sakurajosui, Setagaya-ku, Tokyo, 156-8550, Japan

19 <sup>4</sup>Faculty of Health and Sport Sciences, University of Tsukuba, 1-1-1 Tennodai, Tsukuba-  
20 shi, Ibaraki, 305-8577, Japan

21 <sup>5</sup>Japan Society for the Promotion of Science, 5-3-1 Kojimachi, Chiyoda-ku, Tokyo, 102-  
22 0083, Japan

23 <sup>6</sup>Faculty of Health and Sports Sciences, Toyo University, 1-7-11 Akabanedai, Kita-ku,  
24 Tokyo, 115-8650, Japan

25 <sup>7</sup>Faculty of Health Care and Medical Sports, Teikyo Heisei University, 4-1 Uruidominami,  
26 Ichihara-shi, Chiba, 290-0193, Japan

27 #Co-first author

28

29 **\*Corresponding author:** Kazuhiro Shimizu, PhD.

30 Postal address: Japan Institute of Sports Sciences

31 3-15-1 Nishigaoka, Kita-ku, Tokyo 115-0056, Japan

32 E-mail address: kazuhiro.shimizu@jpnsport.go.jp

33 Telephone: +81 3 5963 0231

34 Fax: +81 3 5963 0232

35

36 **Number of tables:** 4

37 **Number of figures: 4**

38

39 **ABSTRACT**

40 Despite the known health impacts of rapid weight loss (RWL) in athletes,  
41 effective mitigation strategies, especially nutritional ones, remain underexplored. This  
42 study aimed to investigate the effects of *Lactiplantibacillus pentosus* ONRICb0240-  
43 containing food (b240) intake on salivary secretory immunoglobulin A (SIgA) secretion  
44 in judo athletes undergoing RWL. Therefore, 17 male competitive college judo athletes  
45 were categorized into two groups: the b240 intake (active group; n = 9) and no b240  
46 intake (control group; n = 8). Both groups engaged in 4 weeks of regular training  
47 followed by 1 week of RWL. The active group consumed b240 daily, every evening  
48 before dinner, throughout the 5-week experiment. Participants documented upper-  
49 respiratory symptoms (URS) and abdominal conditions during the study. Saliva samples  
50 were collected, and physical fitness tests were performed on each participant at 0 weeks  
51 (before intervention), 4 weeks (before weight loss), and 5 weeks (after weight loss).  
52 Results showed that salivary SIgA secretion was significantly decreased in the control  
53 group after weight loss compared to before the intervention (0 weeks,  $p < 0.05$ ), while  
54 no significant changes were observed in the active group. Compared to the control

55 group, URS decreased, and the percentage of days participants reported their abdominal  
56 condition as “good” increased in the active group ( $p < 0.05$ ). These findings suggest that  
57 regular intake of b240 may be beneficial for reducing the frequency of URS among judo  
58 athletes, potentially due to better maintenance of salivary SIgA secretion during training  
59 and RWL.

60

61 **Key words:** *Lactiplantibacillus pentosus*, judo, salivary SIgA, mucosal immunity, upper-  
62 respiratory symptoms

63 タイトル：

64 柔道選手における急速減量による免疫低下に対する *Lactiplantibacillus pentosus*

65 ONRICb0240 の効果

66 著者名：

67 清水和弘<sup>1,\*</sup>, 服部聡士<sup>2,#</sup>, 松本恵<sup>3</sup>, 平岡拓晃<sup>4</sup>, 田名辺陽子<sup>1,5,6</sup>, 花岡裕吉<sup>1</sup>, 小野

68 卓志<sup>4,7</sup>, 木村文律<sup>4</sup>, 濱田広一郎<sup>2</sup>, 渡部厚一<sup>4</sup>, 久木留毅<sup>1</sup>

69

70 所属機関:

71 <sup>1</sup>115-0056 東京都北区西が丘 3-15-1, 国立スポーツ科学センタースポーツ科学

72 研究部門

73 <sup>2</sup>842-0195 佐賀県神埼郡吉野ヶ里町大曲字東山 5006-5, 大塚製薬株式会社佐賀

74 栄養製品研究所

75 <sup>3</sup>156-8550 東京都世田谷区桜上水 3-25-40, 日本大学文理学部体育学科

76 <sup>4</sup>305-8577 茨城県つくば市天王台1-1-1, 筑波大学体育系

77 <sup>5</sup>102-0083 東京都千代田区麴町5-3-1, 日本学術振興会

78 <sup>6</sup>115-8650 東京都北区赤羽台1-7-11, 東洋大学健康スポーツ科学研究科

79 7290-0193 千葉県市原市うるいど南 4-1, 帝京平成大学健康医療スポーツ学部医療

80 スポーツ学科

81 #Co-first author

82

83 要約

84 アスリートにおける急速減量 (RWL) の健康への影響が知られているにも関わ

85 らず、効果的な軽減策、とくに栄養による解決策は不明である。本研究は、

86 RWL を行う柔道選手の唾液分泌型免疫グロブリン A (SIgA) に対する乳酸菌

87 (*Lactiplantibacillus pentosus* ONRICb0240) 含有食品 (b240) の効果を検討する

88 ことを目的とした。男子大学柔道選手 17 名を対象とし、乳酸菌摂取群 (active

89 群; n = 9) と摂取なし群 (control 群; n = 8) の 2 群に分けた。両群とも 4 週間の

90 通常トレーニングの後に 1 週間の RWL を行った。active 群は 5 週間の実験期間

91 中、毎日夕食前に乳酸菌を摂取した。参加者は試験期間中、上気道症状 (URS)

92 と腹部の状態を記録した。0 週 (介入前)、4 週 (減量前)、5 週 (減量後) に

93 唾液採取と体力テストを行った。その結果、減量後の control 群では、介入前

94 (0 週、 $p < 0.05$ ) と比較して唾液中の SIgA 分泌量が有意に減少したが、active

95 群では有意な変化は認められなかった。また、Control 群に比べて active 群では

96 URS が少なく、腹部の状態について「良好」と回答した日数の割合が多かった

97 ( $p < 0.05$ )。これらの結果より、乳酸菌 b240 の継続的な摂取は減量期の柔道  
98 選手の免疫低下抑制や URS の予防に働き、良好なコンディション維持に役立つ  
99 可能性が示唆された。

100

## 101 INTRODUCTION

102 Judo, an Olympic sport played worldwide, employs a weight class system to ensure  
103 fairness by accounting for individual differences in physical characteristics <sup>1)</sup>. Many  
104 athletes strategically use weight loss to gain a physical advantage over their opponents in  
105 competitions <sup>2)</sup>. Rapid weight loss (RWL) <sup>3)</sup> has been shown to impose significant  
106 physiological and psychological burdens, as detailed in a systematic review <sup>4)</sup>.  
107 Specifically, methods of RWL, such as dietary restriction and dehydration, can cause  
108 various physiological issues, including upper-respiratory symptoms (URS) <sup>5)</sup> and  
109 abdominal problems <sup>6)</sup>.

110 The frequent occurrence of URS is attributed to impaired immune function. Indeed,  
111 athletes undergoing RWL have been observed to exhibit URS alongside reductions in  
112 secretory immunoglobulin A (SIgA) in saliva <sup>7)</sup> and blood lymphocytes <sup>8)</sup>. Furthermore,  
113 intense exercise training has been shown to significantly reduce salivary SIgA secretion  
114 in combat sports <sup>9)</sup>. Therefore, preventing immunosuppression is inferred to be the most

115 effective intervention for inhibiting URS during RWL. Therefore, it is speculated that  
116 preventing immunosuppression is the most effective intervention for suppressing URS in  
117 the RWL.

118 *Lactiplantibacillus pentosus* ONRICb0240, an anaerobic/non-spore-forming gram-  
119 positive bacterium isolated from fermented tea leaves <sup>10)</sup>, secretes SIgA in Peyer's patch  
120 cells in a strain-specific manner <sup>11)</sup>. We hypothesized that its action might contribute to  
121 maintaining immune function in humans. Indeed, it has been demonstrated that heat-  
122 killed ONRICb0240 promotes salivary SIgA secretion in elderly adults with low physical  
123 fitness <sup>12)</sup>. Therefore, intake of heat-killed ONRICb0240 is expected to preserve immune  
124 functions and suppress URS.

125 Athletes who routinely perform intense exercise training often experience abdominal  
126 issues, such as nausea, abdominal cramping, and diarrhea. These problems are believed  
127 to result from a decrease in intestinal membrane permeability, inducing a pro-  
128 inflammatory cascade <sup>6,13)</sup>. This effect is speculated to be more severe in athletes  
129 undergoing RWL because the combination of intense exercise, dehydration (via water  
130 restriction and sweating), and caloric restriction amplifies the elevation of inflammatory  
131 cytokine levels <sup>14)</sup>. Conversely, several clinical trials have indicated that heat-killed  
132 bacteria, such as *Lactiplantibacillus plantarum* L-137 <sup>15)</sup> and *Enterococcus faecalis* EC-



133 12<sup>16)</sup> can regulate the intestinal environment by producing regenerating family member  
134 3 (Reg3), a major antimicrobial peptide. Similarly, heat-killed ONRICb0240 might  
135 promote intestinal regulation through antimicrobial actions, as the bacteria activate toll-  
136 like receptor 2 (TLR-2)<sup>17)</sup>, a receptor upstream of increased Reg3 gene expression<sup>18)</sup>.  
137 Therefore, ONRICb0240 may help inhibit gastrointestinal distress in athletes during  
138 RWL.

139 This study aimed to investigate the effects of ONRICb0240 intake on salivary SIgA  
140 secretion, URS, and abdominal condition in judo athletes undergoing RWL. It is thought  
141 that the ingested b240 is recognized by dendritic cells in the intestinal tract, which  
142 produce cytokines such as interleukin 6 (IL-6) to induce IgA production by B cells. B  
143 cells then contribute to SIgA secretion in the intestinal tract, and some B cells migrate to  
144 the salivary glands and contribute to salivary SIgA secretion<sup>17,19)</sup>. Therefore, we  
145 hypothesized that continuous intake of b240 may reduce the risk of infection and the  
146 occurrence of URS in athletes during RWL by stimulating salivary SIgA secretion and  
147 that its antibacterial effect in the intestinal tract may suppress the deterioration of  
148 abdominal conditions.

149

150 **MATERIALS AND METHODS**

## 151           **Experimental Approach to the Problem**

152           In this study, we investigated the effects of continuous b240 intake on immune function  
153   by requesting judo athletes perform RWL and using salivary SIgA secretion, URS  
154   appearance, and abdominal condition as evaluation indices. This study was conducted in  
155   an open-label randomized controlled trial.

156

## 157           **Participants**

158           This study adhered to the principles of the Declaration of Helsinki and was approved  
159   by the Ethics Committees of Japan Institute of Sports Sciences [approval #033 (2017)].  
160   Participants were provided with detailed information about the study's risks, stressors,  
161   and potential benefits before signing an informed consent form. A total of 26 competitive  
162   male college judo athletes participated in this study (Fig. 1). According to a self-reported  
163   questionnaire, none of the participants had been treated with medications known to affect  
164   immune function, experienced acute infectious illnesses in the preceding 3 months, or  
165   regularly smoked tobacco. Participants were randomly assigned to either the b240 intake  
166   (active group; n = 13) or no b240 intake (control group; n = 13) based on age, height,  
167   body weight, susceptibility to URS, and salivary SIgA secretion rate. Finally, 17  
168   participants (active group: n = 9; control group: n = 8) were included in the analysis (Fig.

169 1). Participants had an average judo practice duration of  $13.1 \pm 0.6$  years. All participants  
170 trained regularly for 16.5 h per week and held technical levels between 1st and 2nd Dan  
171 black belts, competing in weight categories ranging from 66 to 100 kg.

172 [*Insert Fig. 1*]

173

#### 174 **Procedures**

175 The experimental design of this study is illustrated in Fig. 2. Over a 5-week  
176 intervention period, participants were instructed to continue their normal training routines.  
177 In the last week, participants engaged in self-determined weight loss programs, including  
178 dietary energy restriction, fluid restriction, wearing sauna suits during intense exercise  
179 training, and sauna use, in addition to their regular judo training. No food other than that,  
180 including b240 was provided during the experimental periods, and each participant used  
181 his own selection for the intake of each nutrient, etc. The consumption of supplements  
182 (e.g., vitamins, minerals) and additional probiotics (e.g., yogurt) was prohibited during  
183 the study. The active group consumed heat-killed b240 ( $2 \times 10^9$  cells containing) every  
184 evening before dinner for 5 weeks. A trial tablet containing heat-killed ONRICb0240 was  
185 prepared as follows: after cultivating the bacteria and performing repeated washes, the  
186 cells were sterilized in an autoclave and freeze-dried. The non-viable b240 was counted

187 using a flow cytometer and adjusted to produce the test sample with  $2 \times 10^9$  cells. The  
188 nutritional content per 100 g of the food included: energy, 95 kcal; protein, 10 g (L-valine,  
189 500 mg; L-leucine, 1,000 mg; L-isoleucine, 500 mg; L-arginine, 500 mg; whey protein,  
190 7.5 g); carbohydrate, 14 g; fat, 0 g; sodium chloride equivalent, 0.16 g; vitamin B<sub>6</sub>, 5.0  
191 mg; vitamin D, 10.0 µg; citric acid, 1,250 mg. Assessments, including anthropometric  
192 measurements, saliva sampling, subjective physical condition reporting, and physical  
193 fitness tests, were conducted at 7:00 AM at their normal weight at three time points: 0  
194 weeks (before intervention), 4 weeks (before weight loss), and 5 weeks (after weight loss).  
195 During the initial 4 weeks, participants performed their regular judo, interval, and  
196 resistance training regimens for 2.5–3.0 h daily. Judo training sessions included stretching,  
197 judo-specific skills and drills, and high-intensity randori (fighting practice), targeting  
198 approximately 60–80% of the maximal oxygen uptake and 80–85% of the maximum heart  
199 rate <sup>20</sup>.

200

201 *[Insert Fig. 2]*

202

203 **Subjective physical assessments**

204 Participants were asked to evaluate their physical condition over the past week at the  
205 0-, 4-, and 5-week marks using a questionnaire. This questionnaire inquired about the  
206 appearance of URS and the rating of abdominal condition, as described in a previous  
207 report <sup>21)</sup>. The assessment of URS was based on prior studies <sup>22)</sup>, in which complaints of  
208 URS on any day, assuming the presence of at least one URS (sore throat, runny nose,  
209 coughing, headache, lassitude, chillness, and fever, were noted. The percentage of days  
210 with complaints was calculated for each period (before intervention (1 week before the  
211 experiment), normal training (4 weeks), and weight loss (1 week) and compared between  
212 groups. Responses to abdominal conditions (“good”, “usual”, “bad”) were also compared  
213 between groups by calculating the percentage of each response for each period. The data  
214 are presented as the percentage of days with reported conditions for each period.

215

### 216 **Anthropometric measurements**

217 Body weight, percentage of fat, fat mass, fat-free mass, and body water content were  
218 recorded using a multi-frequency bioelectrical impedance device (InBody® 770; InBody  
219 Japan Inc., Tokyo, Japan). Measurements were taken with each participant in light  
220 clothing and without footwear.

221

## 222           **Saliva analyses**

223           Participants were instructed to abstain from alcohol on the day before the test and fast  
224           from 10:00 PM the previous day. Saliva samples were collected between 6:00 AM and  
225           7:00 AM after overnight fasting. Participants rinsed their oral cavity three times with  
226           mineral water and then rested for at least 5 min. Saliva production was stimulated by  
227           chewing paraffin wax at a rate of 180 times/180 s. The stimulated saliva was collected by  
228           passive dribbling into a sterile collection tube. Following the measurement of sample  
229           volume, saliva samples were frozen at -40°C. SIgA concentration was determined using  
230           an enzyme-linked immunosorbent assay <sup>12</sup>). The SIgA concentration ( $\mu\text{g/mL}$ ) was  
231           multiplied by the saliva flow rate over 3 min ( $\text{mL}/3 \text{ min}$ ) to calculate the SIgA secretion  
232           rate ( $\mu\text{g}/3 \text{ min}$ ).

233

## 234           **Physical fitness tests**

235           The participants underwent physical fitness tests at 0, 4, and 5 weeks, encompassing  
236           five tests: isometric grip strength measured with a handgrip dynamometer, muscle  
237           endurance assessed through a 30-s sit-up test, muscular power evaluated using vertical  
238           jump and standing broad jump tests, and whole-body reaction time. The isometric grip  
239           strength, sit-up, and standing broad jump tests followed protocols described in the

240 “Physical Fitness Test” by the Japan Ministry of Education, Culture, Sports, Science, and  
241 Technology. The vertical jump test was performed on a mat switch (660 × 1000 mm)  
242 connected to a PC via an A/D converter (Multi Jump Tester, DKH Co., Ltd., Tokyo,  
243 Japan). Whole-body reaction time was measured by instructing participants to jump  
244 vertically as quickly as possible upon the activation of a red light placed 2 m ahead, with  
245 the time from light stimulus to both feet leaving the ground recorded (T.K.K.1264p, Takei  
246 Scientific Instruments Co., Ltd., Nigata, Japan).

247

#### 248 **Assessment of nutritional intake**

249 Nutritional intake data were collected using a detailed 3-day food record immediately  
250 before the intervention and during the weight loss period. Participants were encouraged  
251 to accurately record all foods and fluids consumed, ensuring precise reporting of amounts  
252 and types. Daily nutritional intake values were calculated using dietary assessment  
253 software (Excel Nutrition ver. 2.3; Kenpakusha, Tokyo, Japan). Participants were also  
254 asked to adhere to their normal dietary patterns throughout the study.

255

#### 256 **Statistical analysis**

257 Data are presented as means  $\pm$  standard error (SE). A  $p$ -value of  $< 0.05$  was considered  
258 statistically significant in all analyses. Variables obtained from questionnaires (including  
259 physical assessments of URS and abdominal conditions) were analyzed using the chi-  
260 squared test and residual analysis. Other variables were analyzed using two-way repeated  
261 measures analysis of variance (ANOVA). A Tukey-Kramer post-hoc test was applied to  
262 identify specific differences when ANOVA revealed significant effects.

263

## 264 **RESULTS**

265 Table 1 presents the physical characteristics of the active and control groups during the  
266 study periods. No significant group-time interaction was found for body weight, fat mass,  
267 fat-free mass, and body water. A significant decrease in body weight (3.6% average loss),  
268 fat mass, fat-free mass, and body water was observed in both groups after weight loss (at  
269 5 weeks) compared to before the intervention (at 0 weeks,  $p < 0.05$ ). The percentage of  
270 body fat in the control group at 5 weeks was significantly decreased compared to that at  
271 0 weeks ( $p < 0.05$ ). Similarly, body weight, fat-free mass, and body water in both groups  
272 at 5 weeks were significantly decreased compared to those before weight loss (at 4 weeks,  
273  $p < 0.05$ ). Fat mass in the control group at 5 weeks was significantly decreased compared  
274 to that at 4 weeks ( $p < 0.05$ ).



275

276 [Insert Table 1]

277

278 Table 2 and Fig. 3 detail salivary SIgA-related measurements. No significant group-  
279 time interaction was found for saliva flow, SIgA concentration, and SIgA secretion rate.  
280 The saliva flow rate at 5 weeks in both groups was significantly decreased compared to  
281 that at 0 weeks ( $p < 0.05$ ), and in the active group, it was significantly decreased compared  
282 to that at 4 weeks ( $p < 0.05$ ). The salivary SIgA secretion rate in the control group at 5  
283 weeks was significantly decreased compared to that at 0 weeks ( $p < 0.05$ ), whereas the  
284 active group did not show a significant change.

285

286 [Insert Table 2]

287 [Insert Fig. 3]

288

289 Fig. 4 illustrates subjective physical assessments, including URS and abdominal  
290 condition. The percentage of days with complaints of URS during the intervention periods  
291 tended to be lower in the active group than in the control group, particularly during the  
292 normal training period, where it was significantly lower in the active group ( $p < 0.05$ ).

293 The percentage of respondents reporting their abdominal condition as “good” in the active  
294 group was significantly increased during the intervention periods compared to that in the  
295 control group ( $p < 0.05$ ).

296

297 *[Insert Fig. 4]*

298

299 Table 3 shows the results of physical fitness tests during the experimental periods. No  
300 significant group-time interaction was found for any fitness test items. Sit-up  
301 performance in both groups at 5 weeks was significantly increased compared to that at 0  
302 weeks ( $p < 0.05$ ). The vertical jump in the control group at 4 weeks was significantly  
303 decreased compared to that at 0 weeks ( $p < 0.05$ ).

304

305 *[Insert Table 3]*

306

307 Table 4 presents nutritional intake during the experimental periods. No significant  
308 group-time interaction was found for any nutritional intake. Energy, carbohydrate, and  
309 water intake in both groups at 5 weeks were significantly decreased compared to those at

310 0 weeks ( $p < 0.05$ ). Protein and fat intake in the active group at 5 weeks were significantly  
311 decreased compared to those at 0 weeks ( $p < 0.05$ ).

312

313 *[Insert Table 4]*

314

## 315 **DISCUSSION**

316 Athletes competing in weight classifications often suffer from URS and various  
317 abdominal problems due to the stress of RWL. Therefore, strategies are needed to  
318 maintain physical condition during competitive events. In this study, we investigated the  
319 effects of b240 on immune function, URS, and abdominal condition in judo athletes  
320 during RWL. Our results showed that approximately 3.6% body weight loss induced  
321 changes in salivary SIgA secretion, although physical fitness levels remained unchanged.  
322 Importantly, b240 intake mitigated the significant decrease in salivary SIgA secretion and  
323 reduced the percentage of URS-affected days during RWL. Additionally, the percentage  
324 of days participants reported having a “good” abdominal condition increased. These  
325 findings indicate that regular ingestion of b240 is beneficial for maintaining a favorable  
326 physical condition in athletes during RWL through the inhibition of immune dysfunction  
327 and the prevention of URS and gastrointestinal distress.

328 Salivary SIgA plays a crucial role as an initial defense mechanism in the human  
329 organism, specifically capturing various pathogens (bacteria and viruses) in the oral  
330 cavity and either inactivating or aggregating them. It has been revealed that salivary SIgA  
331 levels dramatically decrease in athletes under physiological stressors such as long-  
332 distance running <sup>23)</sup>, amenorrhea <sup>24)</sup>, and RWL <sup>5)</sup>. A review examining changes in salivary  
333 SIgA secretion over time in patients with URS found a significant decrease in salivary  
334 SIgA secretion beginning 2-3 weeks before the onset of symptoms <sup>25)</sup>. Therefore,  
335 maintaining salivary SIgA levels several weeks prior to competition is crucial for  
336 ensuring optimal physical condition during the event. In this study, we observed that  
337 RWL reduced salivary SIgA secretion in the control group over the 5-week experimental  
338 period, whereas the active group, who regularly took b240, maintained salivary SIgA  
339 secretion levels. Moreover, URS incidents in the active group were also lower than in the  
340 control group. Therefore, these results suggest that regular ingestion of b240 may inhibit  
341 immunodeficiency and reduce URS over several weeks. Ingested b240 is thought to  
342 activate dendritic cells in the intestinal tract to produce cytokines and induce IgA  
343 production in B cells, which then migrate to the salivary glands and contribute to salivary  
344 SIgA secretion, potentially reducing the risk of infection <sup>17,19)</sup>.

345 The abdominal discomfort in athletes is attributed to an inflammatory cascade, which  
346 increases intestinal membrane permeability. This increase allows for the invasion of  
347 bacteria and toxins due to intense exercise<sup>6,13</sup>). Conversely, the intake of b240 resulted in  
348 a higher percentage of “good” responses regarding abdominal condition compared to  
349 cases without intake. We propose that b240 contributes to this favorable abdominal  
350 condition through the secretion of antimicrobial peptides, such as SIgA, in the intestinal  
351 tract. Certain heat-killed bacteria are known to regulate intestinal function via the  
352 secretion of antimicrobial peptides<sup>15,16</sup>). Moreover, b240 might also improve abdominal  
353 conditions because these heat-killed bacteria stimulate receptors in the intestinal cells,  
354 promoting SIgA secretion<sup>17</sup>). The consumption of *Lactiplantibacillus* has been reported  
355 to positively affect the intestinal environment of athletes<sup>26</sup>), and the results of our study  
356 align with these findings. Notably, the percentage of “good” responses increased not only  
357 during the weight loss phase but also during the normal training phase. Therefore, we  
358 infer that b240 attenuates the excessive increase in intestinal membrane permeability,  
359 contributing to better abdominal conditions in athletes.

360 RWL may exert a greater strain on immune function in a shorter period than exercises  
361 that induce a reduction in SIgA, as previously reported. For instance, peak training over  
362 4 weeks did not affect salivary SIgA concentration and secretion rate in triathletes<sup>27</sup>), and

363 intense training for 21 days also had no effect on cyclists<sup>28)</sup>. Moreover, in a study on  
364 triathletes, it took 20 days for a decrease in salivary SIgA secretion to occur if they did  
365 not maintain sufficient energy intake from their diet<sup>21)</sup>. Therefore, RWL may pose a  
366 higher risk of inducing immune dysfunction than the training conditions previously  
367 reported, at least in terms of salivary SIgA.

368 While we expect that b240 is primarily responsible for the observed improvements in  
369 abdominal condition and immune function, we cannot exclude the possibility that other  
370 components of the intake might have played a role in the study's outcomes. For instance,  
371 it has been suggested that protein intake can inhibit exercise-induced intestinal protein  
372 catabolism<sup>29)</sup> and bacterial/viral infections<sup>30)</sup>. However, the participants in this study  
373 consumed a lower amount of protein than that previously identified as effective in  
374 suppressing these symptoms<sup>31)</sup>. The protein intake of our study participants (active  
375 group: 61.2 g/day, control group: 71.6 g/day, Table 4) was lower than the high protein  
376 intake (220 g/day) reported to be effective<sup>31)</sup>. This discrepancy suggests that abdominal  
377 conditions may not improve without sufficient total protein intake, even with protein  
378 supplementation. Therefore, it appears that b240, rather than the protein content in the  
379 food, notably contributed to the better abdominal condition observed in the active group  
380 compared to the control group, despite the active group's total protein intake being

381 significantly lower during the weight loss period in our study. In addition, while a  
382 relationship between vitamin D and immune function has been established <sup>32,33</sup>), the  
383 vitamin D content in the food (10 µg, approximately 400 IU) in this study was likely  
384 insufficient to impact immune function and URS. Effective vitamin D intake for  
385 increasing salivary SIgA secretion and inhibiting URS is estimated to be approximately  
386 1000–5000 IU <sup>34,35</sup>). This is consistent with previous research indicating that 10 µg of  
387 vitamin D intake in recreational athletes did not affect URS or salivary SIgA secretion <sup>36</sup>).  
388 These findings suggest that the improvements in abdominal condition and reduction in  
389 URS observed in the present study are attributable to the effects of b240.

390 This study had some limitations. First, we prioritized the reduction of salivary SIgA  
391 secretion by approximately 4% during 1 week, based on a previous study <sup>7</sup>), and  
392 allowed each subject to select an RWL method that was easily implementable for them.  
393 Therefore, this study lacked a standardized method for RWL due to variations in b240  
394 intake, training volume, and dehydration methods during the experimental periods. To  
395 achieve the targeted 4% weight loss, participants chose methods that were most feasible  
396 for them. Therefore, this study cannot dismiss the potential influence of individual  
397 RWL methods on the results. Second, the design of this study did not include a placebo  
398 group. Future studies need to demonstrate the effects of ONRICb0240 on immune  
399 function, URS, and abdominal condition in athletes through a double-blind, placebo-  
400 controlled design. Third, the small sample size limits the degree to which the study  
401 results can be generalized. We examined the effects of b240 intake on SIgA secretion in

402 healthy male athletes during the RWL, which presented the difficulty of finding athletes  
403 who played the same sport, had similar years of experience, and conducted RWL.  
404 Additional studies must involve an appropriate number of participants. Fourth, in this  
405 study, the percentage of days with URS complaints over the 4 weeks was significantly  
406 lower in the active group than in the control group; however, no significant changes  
407 were observed in salivary SIgA in either group at 4 weeks. This result may be due to the  
408 intake of b240 affecting not only salivary SIgA but also other immunological factors. It  
409 is thought that ingested b240 is recognized by dendritic cells, which then produce IL-6  
410 to induce IgA production by B cells<sup>17, 19)</sup>. IL-6 induces the activation of T cells,  
411 monocytes, neutrophils, and other cells, in addition to B cells. Therefore, it is possible  
412 that b240 activated these immune cells. In addition, when mice were administered b240  
413 for 2 weeks and subsequently infected with pneumococcus, the production of  
414 inflammatory cytokines in the lungs was reduced compared to that in mice administered  
415 saline<sup>37)</sup>. Furthermore, it has been reported that 5 weeks of b240 intake increased the  
416 expression of the antiviral protein radical S-adenosyl methionine domain containing 2 in  
417 the lungs of mice<sup>38)</sup>. These factors, in addition to SIgA, may contribute to the changes  
418 in URS. However, there is a lack of evidence in humans, and these factors were not  
419 measured in this study; therefore, this remains speculative. Future studies are required  
420 to clarify the effects of b240 intake on these immunological factors in response to  
421 intensive exercise and RWL.

422 In conclusion, we found that RWL could rapidly decrease salivary SIgA secretion  
423 over approximately one week. However, continuous intake of b240 for 5 weeks was



424 found to inhibit reductions in salivary SIgA secretion, the appearance of URS, and  
425 improve abdominal conditions. These results suggest that b240 plays a crucial role in  
426 maintaining physical condition by suppressing the occurrence of URS through  
427 immunosuppression inhibition and enhancing abdominal conditions in athletes during  
428 RWL.

429

#### 430 **Acknowledgements**

431 We thank all the participants for their time and effort in participating in this study. This  
432 study was supported by the JSC Japan High Performance Sport Center Total Conditioning  
433 Research Project and Grants-in-Aid for Scientific Research from the Japan Society for  
434 the Promotion of Science (JP17K01887 to K. S.).

435

#### 436 **Author Contributions**

437 K.S. and S.H conceived the study, the main conceptual ideas, and the proof outline.  
438 M.M. and H.H. support designed in the experimental protocol and setup of the  
439 experimental environment. K.N., Y.T., Y.H., T.O., and F.K. collected the data. K.W.  
440 and K.H. aided in interpreting the results. T.K. supervised the project and advised on the  
441 research concept. All authors discussed the results and commented on the manuscript.

442 **Conflict of interest**

443 Satoshi Hattori and Koichiro Hamada are employees of Otsuka Pharmaceutical Co., Ltd.

444 The other authors have no conflicts of interest to declare.

445

446 **REFERENCES**

- 447 1) Reale R, Slater G and Burke LM. 2017. Acute-weight-loss strategies for combat  
448 sports and applications to Olympic success. *Int J Sports Physiol Perform* 12: 142–151.  
449 doi: 10.1123/ijsp.2016-0211.
- 450 2) Artioli GG, Gualano B, Franchini E, Scagliusi FB, Takesian M, Fuchs M and  
451 Lancha AH, Jr. 2010. Prevalence, magnitude, and methods of rapid weight loss among  
452 judo competitors. *Med Sci Sports Exerc* 42: 436–442. doi:  
453 10.1249/MSS.0b013e3181ba8055.
- 454 3) Reale R, Cox GR, Slater G and Burke LM. 2016. Regain in body mass after  
455 weigh-in is linked to success in real life judo competition. *Int J Sport Nutr Exerc Metab*  
456 26: 525–530. doi: 10.1123/ijsnem.2015-0359.
- 457 4) Lakicevic N, Roklicer R, Bianco A, Mani D, Paoli A, Trivic T, Ostojic SM,  
458 Milovancev A, Maksimovic N and Drid P. 2020. Effects of rapid weight loss on judo  
459 athletes: a systematic review. *Nutrients* 12: 1220. doi: 10.3390/nu12051220.
- 460 5) Hiraoka H, Hanaoka Y, Jesmin S, Kimura F, Matsuish Y, Shimizu K and  
461 Watanabe K. 2019. Variation of salivary IgA during weight loss period before a  
462 competition among university judo players. *J Clin Med Res* 11: 798–806. doi:  
463 10.14740/jocmr3998.

- 464 6) de Oliveira EP, Burini RC and Jeukendrup A. 2014. Gastrointestinal complaints  
465 during exercise: prevalence, etiology, and nutritional recommendations. *Sports Med* 44  
466 Suppl 1: S79–S85. doi: 10.1007/s40279-014-0153-2.
- 467 7) Shimizu K, Aizawa K, Suzuki N, Kukidome T, Kimura F, Akama T, Mesaki N  
468 and Kono I. 2007. Evaluation of condition during rapid weight loss using salivary level  
469 of secretory SIgA in elite wrestlers. *Jap J Clin Sports Med* 15: 441–447.
- 470 8) Shimizu K, Aizawa K, Suzuki N, Masuchi K, Okada H, Akimoto T, Mesaki N,  
471 Kono I and Akama T. 2011. Influences of weight loss on monocytes and T-cell  
472 subpopulations in male judo athletes. *J Strength Cond Res* 25: 1943–1950. doi:  
473 10.1519/JSC.0b013e3181e4f9c6.
- 474 9) Tsai ML, Ko MH, Chang CK, Chou KM and Fang SH. 2011. Impact of intense  
475 training and rapid weight changes on salivary parameters in elite female Taekwondo  
476 athletes. *Scand J Med Sci Sports* 21: 758–764. doi: 10.1111/j.1600-0838.2010.01099.x.
- 477 10) Okada S, Daengsubha W, Uchimura T, Ohara N and Kozaki M. 1986. Flora of  
478 lactic acid bacteria in Miang produced in northern Thailand. *J Gen Appl Microbiol* 32:  
479 57–65.
- 480 11) Yamahira S, Toba M, Kishi K and Okamatsu H. 2006. Stimulation of mucosal  
481 immune system by lactic acid bacteria originating in tea. *J J Lactic Acid Bact* 17: 57–66.

- 482 12) Shimizu K, Sato H, Suga Y, Yamahira S, Toba M, Hamuro K, Kakumoto K,  
483 Kohda N, Akama T, Kono I and Kuno S. 2014. The effects of *Lactobacillus pentosus*  
484 strain b240 and appropriate physical training on salivary secretory IgA levels in elderly  
485 adults with low physical fitness: a randomized, double-blind, placebo-controlled trial. *J*  
486 *Clin Biochem Nutr* 54: 61–66. doi: 10.3164/jcbn.13-62.
- 487 13) Chantler S, Griffiths A, Matu J, Davison G, Jones B and Deighton K. 2021. The  
488 Effects of Exercise on Indirect Markers of Gut Damage and Permeability: A Systematic  
489 Review and Meta-analysis. *Sports Med* 51: 113–124. doi: 10.1007/s40279-020-01348-y.
- 490 14) Abdelmalek S, Chtourou H, Souissi N and Tabka Z. 2015. Caloric restriction  
491 effect on proinflammatory cytokines, growth hormone, and steroid hormone  
492 concentrations during exercise in judokas. *Oxid Med Cell Longev* 2015: 809492. doi:  
493 10.1155/2015/809492.
- 494 15) Nakai H, Murosaki S, Yamamoto Y, Furutani M, Matsuoka R and Hirose Y.  
495 2021. Safety and efficacy of using heat-killed *Lactobacillus plantarum* L-137: High-dose  
496 and long-term use effects on immune-related safety and intestinal bacterial flora. *J*  
497 *Immunotoxicol* 18: 127–135. doi: 10.1080/1547691x.2021.1979698.

- 498 16) Terada A, Bukawa W, Kan T and Mitsuoka T. 2004. Effects of the consumption  
499 of heat-killed *Enterococcus faecalis* EC-12 preparation on microbiota and metabolic  
500 activity of the faeces in healthy adults. *Microb Ecol Health Dis* 16: 188–194.
- 501 17) Kotani Y, Kunisawa J, Suzuki Y, Sato I, Saito T, Toba M, Kohda N and Kiyono  
502 H. 2014. Role of *Lactobacillus pentosus* Strain b240 and the Toll-like receptor 2 axis in  
503 Peyer's patch dendritic cell-mediated immunoglobulin A enhancement. *PLoS One* 9:  
504 e91857. doi: 10.1371/journal.pone.0091857.
- 505 18) Dessein R, Gironella M, Vignal C, Peyrin-Biroulet L, Sokol H, Secher T, Lacas-  
506 Gervais S, Gratadoux JJ, Lafont F, Dagorn JC, Ryffel B, Akira S, Langella P, Nùñez G,  
507 Sirard JC, Iovanna J, Simonet M and Chamailard M. 2009. Toll-like receptor 2 is critical  
508 for induction of Reg3 beta expression and intestinal clearance of *Yersinia*  
509 pseudotuberculosis. *Gut* 58: 771–776. doi: 10.1136/gut.2008.168443.
- 510 19) Aase A, Sommerfelt H, Petersen LB, Bolstad M, Cox RJ, Langeland N,  
511 Guttormsen AB, Steinsland H, Skrede S and Brandtzaeg P. 2016. Salivary IgA from the  
512 sublingual compartment as a novel noninvasive proxy for intestinal immune induction.  
513 *Mucosal Immunol* 9: 884–893. doi: 10.1038/mi.2015.107.

- 514 20) Miyamoto T, Oshima Y, Shigematsu R, Bar-Or Y and Miura K. 1994. Intensity  
515 of randori exercise and anaerobic threshold in judo athletes. *Memories of Osaka Kyoiku*  
516 University, Ser IV Education, Psychology, Special Education and Physical Education 42.
- 517 21) Matsumoto M, Satoh K, Kushi H, Hamuro K, Sakurai M, Saito H, Tanaka R,  
518 Saito T, Kohda N and Hamada K. 2021. Salivary immunoglobulin a secretion rate during  
519 peak period conditioning regimens in triathletes. *J Strength Cond Res* 35: 1389–1396.  
520 doi: 10.1519/jsc.0000000000002918.
- 521 22) Fahlman MM and Engels HJ. 2005. Mucosal IgA and URTI in American college  
522 football players: a year longitudinal study. *Med Sci Sports Exerc* 37: 374–380. doi:  
523 10.1249/01.mss.0000155432.67020.88.
- 524 23) Akimoto T, Akama T, Sugiura K, Tatsuno M, Koda Y, Waku T and Kono I.  
525 1998. Alteration of local immunity in the oral cavity after endurance running. *Jpn J Phys*  
526 *Fitness Sports Med* 47: 53–61.
- 527 24) Shimizu K, Suzuki N, Nakamura M, Aizawa K, Imai T, Suzuki S, Eda N,  
528 Hanaoka Y, Nakao K, Suzuki N, Mesaki N, Kono I and Akama T. 2012. Mucosal immune  
529 function comparison between amenorrheic and eumenorrheic distance runners. *J Strength*  
530 *Cond Res* 26: 1402–1406. doi: 10.1519/JSC.0b013e31822e7a6c.

- 531 25) Lehmann MJ, Lormes W, Opitz-Gress A, Steinacker JM, Netzer N, Foster C and  
532 Gastmann U. 1997. Training and overtraining: an overview and experimental results in  
533 endurance sports. *J Sports Med Phys Fitness* 37: 7–17.
- 534 26) Schreiber C, Tamir S, Golan R, Weinstein A and Weinstein Y. 2021. The effect  
535 of probiotic supplementation on performance, inflammatory markers and gastrointestinal  
536 symptoms in elite road cyclists. *J Int Soc Sports Nutr* 18: 36. doi: 10.1186/s12970-021-  
537 00432-6.
- 538 27) Robson-Ansley PJ, Blannin A and Gleeson M. 2007. Elevated plasma  
539 interleukin-6 levels in trained male triathletes following an acute period of intense interval  
540 training. *Eur J Appl Physiol* 99: 353–360. doi: 10.1007/s00421-006-0354-y.
- 541 28) Slivka DR, Hailes WS, Cuddy JS and Ruby BC. 2010. Effects of 21 days of  
542 intensified training on markers of overtraining. *J Strength Cond Res* 24: 2604–2612. doi:  
543 10.1519/JSC.0b013e3181e8a4eb.
- 544 29) Halseth AE, Flakoll PJ, Reed EK, Messina AB, Krishna MG, Lacy DB, Williams  
545 PE and Wasserman DH. 1997. Effect of physical activity and fasting on gut and liver  
546 proteolysis in the dog. *Am J Physiol* 273: E1073–E1082. doi:  
547 10.1152/ajpendo.1997.273.6.E1073.



- 548 30) Flakoll PJ, Judy T, Flinn K, Carr C and Flinn S. 2004. Postexercise protein  
549 supplementation improves health and muscle soreness during basic military training in  
550 Marine recruits. *J Appl Physiol* (1985) 96: 951–956. doi:  
551 10.1152/jappphysiol.00811.2003.
- 552 31) Witard OC, Turner JE, Jackman SR, Kies AK, Jeukendrup AE, Bosch JA and  
553 Tipton KD. 2014. High dietary protein restores overreaching induced impairments in  
554 leukocyte trafficking and reduces the incidence of upper respiratory tract infection in elite  
555 cyclists. *Brain Behav Immun* 39: 211–219. doi: 10.1016/j.bbi.2013.10.002.
- 556 32) Prietl B, Treiber G, Pieber TR and Amrein K. 2013. Vitamin D and immune  
557 function. *Nutrients* 5: 2502–2521. doi: 10.3390/nu5072502.
- 558 33) Dubnov-Raz G, Rinat B, Hemilä H, Choleva L, Cohen AH and Constantini NW.  
559 2015. Vitamin D supplementation and upper respiratory tract infections in adolescent  
560 swimmers: a randomized controlled trial. *Pediatr Exerc Sci* 27: 113–119. doi:  
561 10.1123/pes.2014-0030.
- 562 34) Scott JM, Kazman JB, Palmer J, McClung JP, Gaffney-Stomberg E and Gasier  
563 HG. 2019. Effects of vitamin D supplementation on salivary immune responses during  
564 Marine Corps basic training. *Scand J Med Sci Sports* 29: 1322–1330. doi:  
565 10.1111/sms.13467.

- 566 35) Jung HC, Seo MW, Lee S, Kim SW and Song JK. 2018. Vitamin D<sub>3</sub>  
567 supplementation reduces the symptoms of upper respiratory tract infection during winter  
568 training in vitamin D-insufficient taekwondo athletes: a randomized controlled trial. *Int*  
569 *J Environ Res Public Health* 15: 2003. doi: 10.3390/ijerph15092003.
- 570 36) Da Boit M, Gabriel BM, Gray P and Gray SR. 2015. The Effect of Fish Oil,  
571 Vitamin D and protein on URTI incidence in young active people. *Int J Sports Med* 36:  
572 426–430. doi: 10.1055/s-0034-1394464.
- 573 37) Tanaka A, Seki M, Yamahira S, Noguchi H, Kosai K, Toba M, Morinaga Y,  
574 Miyazaki T, Izumikawa K, Takeya H, Yamamoto Y, Yanagihara K, Tashiro T, Kohda N  
575 and Kohno S. 2011. *Lactobacillus pentosus* strain b240 suppresses pneumonia induced  
576 by *Streptococcus pneumoniae* in mice. *Lett Appl Microbiol* 53: 35–43. doi:  
577 10.1111/j.1472-765X.2011.03079.x.
- 578 38) Kiso M, Takano R, Sakabe S, Katsura H, Shinya K, Uraki R, Watanabe S, Saito  
579 H, Toba M, Kohda N and Kawaoka Y. 2013. Protective efficacy of orally administered,  
580 heat-killed *Lactobacillus pentosus* b240 against influenza A virus. *Sci Rep* 3: 1563. doi:  
581 10.1038/srep01563.
- 582
- 583

584 **Figure Legends**

585 **Fig.. 1.** Flow diagram for study participants

586

587 **Fig.. 2.** Experimental design

588 Subjects are divided into the control group and active group. The study consists of a

589 normal training period (4 weeks) and a weight loss period (1 week), totaling 5 weeks of

590 experimental periods. Subjective physical assessments, anthropometric measurements,

591 physical fitness tests, and saliva sampling are conducted at the time points of 0, 4, and 5

592 weeks.

593

594 **Fig. 3.** Change in Salivary SIgA secretion rate during the study periods

595 Values are presented as means  $\pm$  standard error (SE). \*, values that differ significantly

596 from pre-intervention (Pre) within the group ( $p < 0.05$ ).

597

598 **Fig. 4.** Subjective physical assessments of upper-respiratory symptoms (URS) and

599 abdominal condition. The percentage of days with URS (A) and ratings of abdominal

600 conditions (“good”, “usual”, “bad”) (B) before intervention (0 weeks), after 4 weeks of

601 intervention (4 weeks), and after 1 week of weight loss (5 weeks).

602 Table 1. Physiological characteristics of the active and control groups during the  
 603 study periods

		0 week	4 week	5 week
Age (yr)	Active	20.2 ± 0.4		
	Control	19.5 ± 0.4		
Height (cm)	Active	172.1 ± 1.6		
	Control	173.9 ± 1.7		
Body mass (kg)	Active	79.8 ± 5.8	79.8 ± 5.7	76.5 ± 5.3 *, †
	Control	83.2 ± 4.1	83.7 ± 4.0	79.9 ± 3.9 *, †
Fat (%)	ACT	14.4 ± 1.9	13.9 ± 2.2	14.0 ± 2.1
	CON	16.2 ± 1.8	16.1 ± 1.7	15.4 ± 1.8 *
Fat (kg)	Active	12.3 ± 2.9	12.0 ± 3.1	11.5 ± 2.8 *
	Control	13.9 ± 2.2	13.9 ± 2.2	12.7 ± 2.1 *, †
Fat free mass (kg)	Active	67.5 ± 3.1	67.8 ± 2.9	65.0 ± 2.8 *, †
	Control	69.3 ± 2.4	69.9 ± 2.4	67.2 ± 2.3 *, †
Body water (kg)	Active	49.6 ± 2.3	49.8 ± 2.2	47.6 ± 2.1 *, †
	Control	50.8 ± 1.7	51.2 ± 1.7	49.2 ± 1.7 *, †

604

605

606

607

608

Values are presented as means ± SE. \*Values that significantly differ from those at 0 week within the same group ( $p < 0.05$ ). †Values that significantly differ from those at 4 week within the same group ( $p < 0.05$ ).

609 Table 2. Saliva flow rate and salivary secretory immunoglobulin A concentration  
 610 for the active and control groups during the study periods  
 611

		0 week	4 week	5 week
Saliva flow rate (ml/3min)	Active	4.30 ± 0.65	4.61 ± 0.58	2.79 ± 0.41 *, †
	Control	4.45 ± 0.60	4.03 ± 0.64	3.31 ± 0.68 *
SIgA concentration (µg/ml)	Active	73.3 ± 8.1	74.2 ± 10.3	92.6 ± 20.0
	Control	65.9 ± 6.0	62.0 ± 5.4	58.7 ± 7.9

612 Values are presented as means ± SE. \*Values that significantly differ from those  
 613 at 0 week within the same group ( $p < 0.05$ ). †Values that significantly differ from those  
 614 at 4 week within the same group ( $p < 0.05$ ).  
 615  
 616  
 617

618 Table 3. Physical fitness test results for the active and control groups during the  
 619 study periods

		0 week	4 week	5 week
Grip strength (kg)	Active	46.3 ± 2.8	46.8 ± 2.8	46.5 ± 2.8
	Control	45.4 ± 2.1	44.7 ± 2.1	45.1 ± 2.0
Sit-up (times/30sec)	Active	34.5 ± 1.2	35.9 ± 1.1	37.3 ± 1.3 *
	Control	36.8 ± 3.3	37.8 ± 3.2	40.3 ± 2.8 *
Vertical jump (cm)	Active	43.4 ± 2.9	42.4 ± 2.7	43.6 ± 2.7
	Control	42.1 ± 1.8	39.3 ± 2.1 *	41.8 ± 2.0
Standing broad jump (cm)	Active	211 ± 10	209 ± 6	212 ± 7
	Control	210 ± 10	220 ± 7	220 ± 8
Whole body reaction time (msec)	Active	0.32 ± 0.01	0.32 ± 0.02	0.32 ± 0.01
	Control	0.30 ± 0.01	0.30 ± 0.01	0.30 ± 0.01

620

621

622 Values are presented as means ± SE. \*Values that significantly differ from those  
 at 0 week within the same group ( $p < 0.05$ ).

623

624

625 Table 4. Nutritional intake of the active (ACT) and control (CON) groups before  
 626 and after 5 weeks of intervention

		0 week	5 week
Energy (kcal/day)	ACT	2,299 ± 135	1,605 ± 123 *
	CON	2,592 ± 75	1,857 ± 123 *
Protein (g/day)	ACT	76.0 ± 3.8	61.2 ± 3.8 *
	CON	87.6 ± 6.2	71.6 ± 9.3
Fat (g/day)	ACT	60.8 ± 2.0	44.7 ± 3.7 *
	CON	70.8 ± 2.4	58.1 ± 5.5
Carbohydrate (g/day)	ACT	345 ± 28	230 ± 24 *
	CON	381 ± 15	250 ± 17 *
Water (g/day)	ACT	883 ± 65	630 ± 78 *
	CON	1,082 ± 56	720 ± 37 *

627

628

629

630

631

632

633

634

Values are presented as means ± SE. \*Values that significantly differ from those at 0 week within the same group ( $p < 0.05$ ).